

PRE-CLINICAL DEVELOPMENT AND CHARACTERIZATION OF A DECITABINE-INDUCED REGULATORY HLAG⁺CD4⁺-T CELL-ENRICHED CELL PRODUCT AGAINST GVHD

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Background Graft-vs-host-disease (GvHD) is a life-threatening complication of allogeneic hematopoietic cell transplantation(allo-HCT) for which the approved therapies are suboptimal. HLA-G is an immunosuppressive molecule playing a central role in preventing the rejection of the semi-allogeneic foetus during pregnancy, the expression of which is epigenetically regulated. We have previously reported the *ex vivo* generation of decitabine-induced HLAG⁺CD4⁺FOXP3⁻ regulatory T-cells exerting potent suppressive function *in vitro* (Cytotherapy 2017;19:521). Herein, we aimed to further elucidate the molecular mechanisms beyond the HLAG⁺CD4⁺ T cell-mediated immunosuppression and produce and characterize a clinical scale product enriched for HLA-G⁺ immunosuppressive cells(iG-Tregs).

Methods Peripheral Blood Mononuclear Cells from healthy donors were enriched for T-cells through monocyte depletion, activated with CD3/CD28-beads for 3 days, followed by a 3-day decitabine treatment(n=10). The final product(iG-Treg) was assessed *in vitro* and *in vivo*(NSG mice). FACS-sorted HLAG⁺CD4⁺ T-cells were further characterized in depth by RNA-seq (Illumina) and single cell RNA-seq(scRNA-seq) (10X Genomics).

Results The iG-Tregs contained median(Δ m) 93,7% (91.6%-98.6%) CD3⁺cells, predominantly CD4⁺[Δ m CD4:CD8 ratio: 2,05(0.81-3.39)]. Compared to untreated controls(PBS), iG-Tregs were enriched for HLAG⁺ cells within the total CD3⁺ population[Δ m 15.32% (6.9-22.9%)] as well as within the CD4⁺[Δ m 21.6% (6.5-28.9%) and CD8⁺[Δ m 16% (8.3-26.1%)] subpopulations. iG-Tregs demonstrated a favorable safety profile over PBS-treated control cells *in vitro*, as depicted by their diminished alloreactivity against allogeneic PHA blasts (1.8 \pm 1% lysis vs 49 \pm 4, p<0.0001) and their reduced ability to react after PMA/ionomycin stimulation(intracellular IL-2, p=0.02, IFN γ , p=0.008, IL-17a, p=0.04]. Except IL-13 (p=0.05) no other Th1/Th2 cytokines were increased in iG-Treg supernatants. The diminished

alloreactivity of iG-Tregs over PBS-treated control cells was shown also *in vivo* after infusion in NSG mice; 6/9(67%) iG-Treg treated mice survived until sacrifice whereas all 7 PBS-control treated mice developed GvHD from day 21 and succumbed in Δ m day 35. The ability of iG-Tregs in preventing GvHD was tested by their coinfusion with T-cells; while T-cell infusion resulted in lethal GvHD by day 36 in all 3 control mice, the co-administration of iG-Tregs delayed (5/6 mice died by day 66) or even prevented(1/6) the GvHD onset. NGS sequencing showed that FACS-sorted decitabine-induced HLAG⁺CD4⁺ cells had a distinct and homogenous expression profile compared to their HLAG⁻CD4⁺ counterparts. HLAG⁺CD4⁺ cells differentially expressed regulatory genes (e.g. CCL17, CCL22 and CXCL9) and interestingly also, the predominantly myeloid suppressor gene IDO-1, a finding which was further validated at mRNA (RT-PCR) and protein (FACS) level. The prominent regulatory signature of the HLAG⁺CD4⁺ cells was confirmed by single cell RNA-seq analysis.

Conclusions iG-Tregs can be easily produced by a GMP-compatible protocol and constitute a well-characterized product with a safety profile to be further evaluated in an upcoming phase I-II clinical trial for the prevention and treatment of steroid-refractory GvHD(EUDRACT 2021-006367-26). Besides HLA-G, the possible contribution of IDO and other regulatory genes in the immunosuppressive function of HLAG⁺CD4⁺ cells remain to be assessed in future functional assays. To further optimize the ability of the iG-Tregs in GvHD prevention *in vivo*, we currently explore a different treatment design, aiming to increase the persistence of the hypomethylation effect exerted by decitabine.